

Synthesis of 2-alkylisoflavones under phase-transfer catalysis conditions

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2-alkyl isoflavones were synthesised with the aliphatic acid chloride and 2-hydroxydeoxybenzoin under phase-transfer catalysis in acetone–K₂CO₃ medium involves the modified Baker–VenKataraman transformation

Keywords: tetrabutylammonium hydrogen sulfate, aliphatic acid chloride, 2-hydroxydeoxybenzoin, 2-alkyl isoflavone

Isoflavones are widely found in a number of natural products. These natural products have demonstrated numerous biological activities such as antioxidant,¹ anti-inflammatory,² antihypertensive and anticarcinogenic activities.³

Previously the synthesis of 2-alkyl isoflavones can be achieved by adopting the Kostanecki–Robinson reaction,^{4,5} the method got the 2-alkyl isoflavone through the reaction of aliphatic acid anhydrides and 2-hydroxydeoxybenzoin in the presence of the sodium or potassium salt of the corresponding acid under the refluxing condition, but the reaction needed high-temperature and long reaction time. Recently the Kostanecki–Robinson reaction was modified, and the sodium salt of the aliphatic acid was replaced with triethylamine,^{6–8} the modification lowered the reaction temperature, but the yield is not very good.

The modified Baker–VenKataraman transformation were extensively used to synthesise the flavone compounds,⁹ and the 2-phenylisoflavones¹⁰ and 2-furylisoflavones¹¹ also were synthesised. The method use 2-hydroxydeoxybenzoin and aromatic acid chloride to react and get isoflavones in acetone–K₂CO₃ medium with refluxing, the procedure was very simple. Recently 2-arylisoflavones were also acquired with high yield under phase transfer catalysis conditions using tetrabutylammonium hydrogen sulfate as phase transfer catalyst, benzene as a solvent and 20% aq. potassium carbonate as a basic catalyst.¹²

Initial attempts to synthesise 7-hydroxyl-2-methylisoflavone with acetyl chloride and 2,4-dihydroxydeoxybenzoin in acetone–K₂CO₃ medium involve the modified Baker–VenKataraman transformation, but did not give the 7-hydroxyl-2-methylisoflavone. When we added tetrabutylammonium hydrogen sulfate, we obtained the 7-hydroxyl-2-methylisoflavone. When the acetyl chloride was 3 equivalents and tetrabutylammonium hydrogen sulfate was 0.4 equivalents, the yield reached 85% (Table 1).

This success led us to study a variety of 2-hydroxydeoxybenzoin and aliphatic acid chlorides. Two new isoflavones: 7-hydroxyl-2-propylisoflavone and 7-hydroxy-2-butylisoflavone were obtained (Table 2).

In summary, we have described a practical, efficient and inexpensive pathway for the synthesis of 2-alkylisoflavones in excellent yields using tetrabutylammonium hydrogen sulfate as phase transfer catalyst in acetone–K₂CO₃ medium involves the modified Baker–VenKataraman transformation. The availability of tetrabutylammonium hydrogen sulfate and the

Table 1 Synthesis of 7-hydroxyl-2-methylisoflavone with the change of amount of acetyl chloride and tetrabutylammonium hydrogen sulfate

No.	Acetyl chloride/ equiv	TBAHSO ₄ / equiv	Reaction time/h	Yield/%
1	0.2	0	10	0
2	0.2	0.01	10	8
3	0.2	0.05	10	15
4	0.2	0.10	10	24
5	0.2	0.20	10	50
6	0.2	0.40	10	65
7	0.2	0.50	10	71
8	0.3	0.40	10	85

mild reaction condition should find a widespread use as the method of choice to synthesise 2-alkylisoflavones.

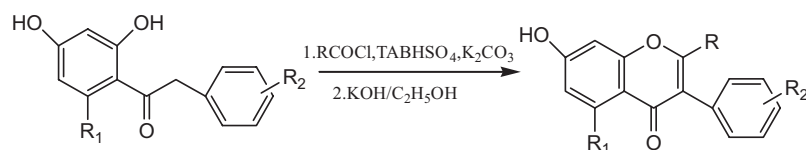
General procedure for synthesising 2-alkyl isoflavones

¹H NMR spectra were measured at 400 MHz on a Varian-Inova 400 spectrometer, ¹³C NMR were measured at 200 MHz on a Bruker–ACE200 spectrometer, and chemical shifts are reported in parts per million (ppm) (δ) relative to TMS as internal standard. Mass spectra (ESI) were determined on API3000 spectrometer and reported as *m/z*. Microanalyses were measured using a Carbo-Erba110G microelemental analyser.

2-hydroxydeoxybenzoin (10 mmol) in acetone (45 ml), tetrabutylammonium hydrogen sulfate (TBAHSO₄, 4 mmol) and anhydrous potassium carbonate (4.5 g) were placed in a flask and aliphatic acid chloride (30 mmol) was added dropwise with stirring during 15–20 min. The mixture was stirred for 2 h at room temperature, then was refluxed for 10 h at 75°C with stirring. The mixture was cooled down and filtered, and solvent of the filter was removed under reduced pressure to give a semi-solid which was refluxed with 5% ethanolic potassium hydroxide (15 ml) for 20 min. The crude semi-solid after removal of ethanol from solution was diluted with water (5 ml) and acidified with dilute hydrochloric acid (15 ml). The resulting solid was filtered, washed with 5% aq. Sodium bicarbonate solution, dried and recrystallised. from the ethanol to give the corresponding isoflavones.

Data for 7-hydroxy-2-propylisoflavone, m.p. 174–176°C, ¹H NMR, d₆-DMSO, δ 0.81–0.84(t, 3H), 1.59–1.69(m, 2H), 2.44–2.52(m, 2H), 6.86–6.864(d, 1H), 6.91–6.93(dd, 1H), 7.23–7.45(m, 5H arom), 7.88–7.90(d, 1H) 10.79(s, 1H, –OH). ¹³C NMR δ: 175.24, 165.17, 162.75, 157.36, 133.63, 130.67, 128.23, 127.25, 127.57, 122.65, 115.78, 115.00, 102.14, 33.66, 20.34, 13.84. *m/z* 280.1 *Anal.* Calcd. for C₁₈H₁₆O₃: C, 77.12; H, 5.75. Found C, 77.18, H, 5.71

Data for 7-hydroxy-2-butylisoflavone, m.p. 162–164°C, ¹H NMR, d₆-DMSO, δ 0.74–0.77(t, 3H), 1.16–1.26(m, 2H), 1.55–1.63(m, 2H), 2.44–2.53(m, 2H), 6.85–6.86(d, 1H), 6.90–6.93(dd, 1H), 7.22–7.45(m, 5H arom), 7.88–7.90(d, 1H), 10.78(s, 1H, –OH). ¹³C NMR



Scheme 1

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Table 2 Synthesis of 2-alkylisoflavone with the change of structure of aliphatic acid chloride and 2-hydroxydeoxybenzoin

Entry no.	R ₁	R ₂	RCOCl	Yield/%	M.p./°C (lit ^{ref})
1	H	H	CH ₃	85	237–238 (240) ⁴
2	H	4-OCH ₃	CH ₃	82	286–287 (286–287) ¹³
3	H	4-OCH ₃	CH ₃ CH ₂	80	242–244 (242–243.5) ¹³
4	H	2-OCH ₃	CH ₃	80	225–227 (225–227) ¹⁴
5	H	2-OCH ₃	CH ₃ CH ₂	76	191–193 (193–194) ⁷
6	OH	H	CH ₃	55	226–228 (228) ⁴
7	OH	4-OCH ₃	CH ₃	60	174–176 (175–176) ¹⁵
8	OH	4-OCH ₃	CH ₃ CH ₂	61	229–231 (229–230) ¹³
9	OH	2-OCH ₃	CH ₃	52	150–151 (150–151) ¹⁵
10	H	H	CH(CH ₃) ₂	60	271–274 (273–275) ⁸
11	H	H	CH ₃ CH ₂ CH ₂	60	174–176
12	H	H	CH ₃ CH ₂ CH ₂ CH ₂	40	162–164

δ: 175.21, 165.45, 162.75, 157.36, 133.61, 130.88, 128.22, 127.26, 127.57, 122.50, 115.79, 114.99, 102.14, 31.62, 29.01, 21.61, 13.57. *m/z* 294.1. *Anal.* Calcd. for C₁₉H₁₈O₃; C, 77.53; H, 6.16. Found C, 77.49; H, 6.17

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References

- R. Haba, S. Watanabe, Y. Arai, H. Chiba and T. Miura, *Env. Health. Prev. Med.*, 2000, **7**, 64.
- H.J. Teede, B. McGrath, L. DeSilva M. Cehun, A. Fassoulakis and P.J. Nestel, *Arterioscler. Thromb. Vasc. Bio*, 2003, **23**, 1066.
- S. Watanabe, S. Uesugi and Y. Kikuchi, *Biomed. Pharmacother.*, 2002, **56**, 302.
- W. Baker and R. Robinson, *J. Chem. Soc.*, 1925, 1981.
- I.M. Heilbron, D.H. Hey and B. Lythgoe, *J. Chem. Soc.*, 1936 295.
- V. Szabó, E. Farkas and A. Lévai, *Acta. Phys. Chim. Debrecina*, 1970, 191.
- A. Levai and T. Patonay, *J. Heterocyclic Chem.*, 2000, **37**, 1065.
- W. Adam, P.B. Rao, H.G. Degen, A. Levai, T. Patonay and C.R. SahaMoller, *J. Org. Chem.*, 2002, **67**, 259.
- A.C. Jain, S.K. Mathur and T.R. Seshadri, *J. Sci. Ind. Res.*, 1962, **21B**, 214.
- V.N. Gupta and T.R. Seshadri, *J. Sci. Ind. Res.*, 1957, **16B**, 116.
- A.V.S. Rao and N.V.S. Rao, *Current Sci.*, 1966, **35**, 149.
- T.U. Kumari, G.L.D. Krupadamam and G. Srimanarayana, *Indian J. Chem.*, 1998, **37B**, 847.
- G.W. Moersch, D.F. Morrow and W.A. Neuklis, *J. Med. Chem.*, 1967, **10**, 154.
- T.R. Seshadri and S. Varadarajan, *Proc. Indian Acad. Sci.*, 1953, **37A**, 784.
- S.P. Ondarenko, A.V. Levenets, M.S. Frasinuk and V.P. Khilya, *Chem. Nat. Compd.*, 2003, **39**, 271.